<u>REVIEW</u>

Osteoarthritis, an Inflammatory Disease

Potential Implication for the Selection of New Therapeutic Targets

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Osteoarthritis (OA) is a well-known disease that is part of the aging process and also one of the most common diseases among mammals. Although this musculoskeletal disorder has been described in mammals of many ages, having been reported in Egyptian mummies and in dinosaurs, its exact etiology is far from being fully understood. With the graying of the world population, it is of the utmost importance to find out more about the pathogenesis of the disease and thus allow the discovery of new treatments to stop or prevent its progression.

A number of risk factors have lately been identified (1). Mechanical factors, among others, are likely to play a very important role in the initiation of the disease process. Endogenous factors such as type II collagen mutation or dysplastic conditions are also known to be involved in initiating the OA process (2).

There is now strong evidence that the structural changes globally observed in OA are due to a combination of factors, ranging from the mechanical to the biochemical (3,4). The disease process affects not only the cartilage, but also the entire joint structure, including the synovial membrane, subchondral bone, ligaments, and periarticular muscles. In OA synovium, the inflammatory changes that take place include synovial hypertrophy and hyperplasia with an increased number of lining cells, and also an infiltration of the sublining tissue with a mixed population of inflammatory cells. In patients with severe disease, the extent of inflammation can sometimes reach that observed in rheumatoid arthritis (RA) patients at the clinical stage (5,6). Some degree of synovitis has also been reported in even the early stages of the disease (7). Synovial inflammation is clearly reflected in many of the signs and symptoms of OA, including joint swelling and effusion, stiffness, and sometimes redness, particularly at the level of the proximal interphalangeal (PIP) and distal interphalangeal (DIP) joints.

Role of inflammation in disease progression: what is the evidence?

The question is whether synovitis in OA is an "innocent bystander" or truly participates in the structural changes of the disease. Moreover, is synovial inflammation only relevant during the "flare" of the disease or is it an ongoing process that permanently contributes to the progression of the disease after it is established? From all observations, there are at least 2 major questions that could be raised regarding synovial inflammation and OA. First, what evidence do we have that inflammation is associated with disease progression? Second, what are the inflammatory factors that could possibly be involved in the genesis of OA structural changes?

The association between OA progression, the signs and symptoms of inflammation, and disease activity has been the subject of a number of interesting studies. One must first recognize that there is still some ambiguity in the definition of disease activity with regard to OA. Some of the criteria used address the functionality of the patient, whereas others, such as stiffness, joint effusion, and other related criteria, probably reflect more accurately the state of joint inflammation. Currently, there are no validated measures of disease activity for OA. The disease progression is commonly measured by change in disease status over a period of time. There are a number of methods available for monitoring

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Marker, disease characteristic, measure	First author	Ref. no.
Cartilage oligomeric protein		
Synovitis, cartilage degradation		
Disease progression	Clark AG	10
	Sharif M	9
	Sharif M	12
Risk factor	Petersson IF	11
Hyaluronic acid		
Synovitis		
Joint score (OA)	Goldberg RL	14
C-reactive protein	5	
Inflammation		
Disease progression	Spector TD	13

Table 1. Examples of reports on the correlation between biologic markers of inflammation and the appearance, progression, and risk factors of osteoarthritis (OA)

disease progression, such as radiography, magnetic resonance imaging (MRI), and arthroscopy, but radiographs are still considered the gold standard. However, radiographs have been proven to have limited sensitivity in the measurement of disease progression. This makes MRI, which provides a more timely and precise structural measurement, a more attractive solution for the future (8).

The main observations that suggest an association between inflammation and the progression of structural changes in OA are derived from clinical studies (9–14). A number of those studies have lately demonstrated an interesting possible association between synovitis, OA inflammation, and progression of structural changes. There are a number of biologic markers that are believed to be associated with OA synovial inflammation, such as cartilage oligomeric protein (COMP), the serum level of C-reactive protein (CRP), and hyaluronic acid (HA) (15–19) (Table 1). It is generally believed that high disease activity suggests a rapid progression of the disease.

COMP is a component of the articular cartilage extracellular matrix and is found in high concentrations in articular cartilage ($\sim 0.1\%$ wet weight). Because this protein is formed by activated synovial cells, it is speculated that elevated COMP levels may reflect synovitis (9–11,20). Other studies (12,13) have shown that elevated CRP levels are predictive of radiographic progression of long-term knee OA. Moreover, it was reported that in women with mild-to-moderately severe knee OA whose disease either progressed or showed no progression, a small elevation in CRP levels was of predictive value (13). The level of another biologic marker, HA, rises in concentration during inflammation (14). HA has been reported to be elevated in OA, and plasma HA levels were found to correlate with an objective functional capacity score and with an articular index based on the total amount of cartilage in the involved joints.

A study by Verbruggen and Veys (21) demonstrated that patients with hand OA, involving the distal joints (DIP, PIP), are generally asymptomatic when the disease is "nonerosive" and become symptomatic during inflammatory episodes. The latter are associated with the onset of erosive OA changes, as seen by sequential roentgenograms.

These data strongly suggest, from both the biologic and clinical sides, an association between joint inflammation and the progression of structural changes in OA.

Role of inflammatory mediators in OA: what is the proof?

Although the roles of inflammation and of inflammatory mediators in the pathophysiology of OA have been under extensive scrutiny in the last decade and a great deal of progress has been made, we do not yet understand all of the ramifications of the systems. Many of the etiologic factors responsible for the initiation of the disease, which happens at the cartilage level and is related to the breakdown of the extracellular macromolecules, remain, however, largely unknown. It is largely agreed that the presence of the synovial inflammation that is often associated with the OA process is believed to be a secondary phenomenon related to the destruction of cartilage and the release of cartilage breakdown products in the synovial fluid. In fact, a number of cartilage macromolecules have been demonstrated to have significant immunogenic properties. For instance, evidence has been provided that OA patients express cellular immunity to the cartilage proteoglycan link protein and C1 domain. Moreover, im-

Figure 1. Schematic diagram of activators (+) and inhibitors (-) of matrix metalloprotease (MMP) synthesis and activity. PA = plasminogen activator; TIMPs = tissue inhibitors of matrix metalloproteases; PKC = protein kinase C; NF- κ B = nuclear factor κ B; MT-MMPs = membrane type matrix metalloproteases.

mune complexes containing antibodies to type II collagen have been detected in the superficial layer of OA cartilage (22). The morphologic changes observed in OA synovium are usually a mild or moderate synovitis that, on occasion, is almost indistinguishable from that in patients with an inflammatory arthritis such as RA (5,6,23,24). This is characterized by an increase in the number of inflammatory mononuclear cells in the sublining tissue, including activated B cells and T lymphocytes (25,26).

Chondrocytes as a source of matrix metalloprotease (MMP) and inflammatory mediator production. There is now strong evidence for major involvement of the MMP family in early cartilage structural changes (3,27). Other enzymes from the serine- and cysteinedependent protease families, such as the plasminogen activator/plasmin system and cathepsin B, respectively, as well as membrane type MMPs also play a role, but primarily as activators of MMPs (Figure 1). Another protease, aggrecanase (a member of the adamalysin family), appears responsible for proteoglycan cleavage as found in human OA synovial fluid.

It is increasingly appreciated that chondrocytes have the capacity to produce a variety of cytokines and mediators associated with inflammation. Chondrocytes obtained from patients with OA actively produce nitric oxide (NO), prostaglandins, interleukin-1 β (IL-1 β), tumor necrosis factor α (TNF α), IL-6, and IL-8. Since these cells are sequestered within the lacunae of articular cartilage (a tissue that is avascular and aneural), the production of such mediators by chondrocytes may not be associated with the classic signs of inflammation. However, as will be discussed, there is evidence that these molecules act within cartilage in an autocrine or paracrine manner to promote a catabolic state, which leads to progressive cartilage damage in OA (28).

Proinflammatory cytokines. It is likely that the excessive production of cytokines and growth factors by the inflamed synovium and activated chondrocytes play an important role in the pathophysiology of OA (3,29). These factors are closely associated with functional alterations not only in the synovium, but also in the cartilage and subchondral bone. They appear to be first produced by the synovial membrane, and diffused into the cartilage through the synovial fluid. They activate the chondrocytes, which, in turn, could produce proteases and other catabolic factors such as NO, and are responsible for inducing cartilage catabolism, chondrocyte apoptosis, and other structural changes associated with the disease process.

A large number of cytokines (pro- and antiinflammatory), antagonists, and growth factors are likely to be involved in OA pathophysiology. Proinflammatory cytokines have been demonstrated to play a pivotal role in the development of this disease process. In particular, IL-1 β and TNF α seem prominent and of major importance to cartilage destruction (30,31). IL-1 β and TNF α can stimulate their own production and induce chondrocytes and synovial cells to produce other cytokines, such as IL-8, IL-6, and leukocyte inhibitory factor (LIF), as well as stimulate proteases and prostaglandin E₂ (PGE₂) production. Moreover, TNF α has also been shown to induce osteoclastic bone resorption in vitro (32), a phenomenon that may be involved in the remodeling of OA subchondral bone.

IL-1 β is primarily synthesized as a precursor, and released in the extracellular milieu in an active form. A protease named IL-1 β -converting enzyme (ICE), or caspase 1, which is located in the plasma membrane, is responsible for generating the mature form of this cytokine (33). The level of this enzyme has been shown to be up-regulated in both OA synovium and cartilage (34). The biologic activation of cells by IL-1 is mediated through an association with specific cell-surface receptors (IL-1R). Two receptors have been identified, type I and type II IL-1R (35). The type I receptor, which has a slightly higher affinity for IL-1 β than for IL-1 α , is responsible for signal transduction. The number of type I IL-1R has been demonstrated to be significantly increased in OA chondrocytes and synovial fibroblasts (36,37), giving these cells a higher sensitivity to stimulation by IL-1 β (36). This phenomenon is responsible for potentiating the effect of this cytokine and up-regulating the gene expression of a number of catabolic factors, which, in turn, enhances cartilage destruction. Both types of IL-1R can also be shed from the cell surface,



and these are named soluble IL-1 receptors (sIL-1R). The shed receptor may function as a cytokine antagonist because the ligand-binding region is preserved. They are believed to act as physiologic inhibitors that regulate the activation of IL-1R. Recent observations indicate that the type II receptor may be several-fold more potent than the type I receptor in antagonizing the catabolic effects of IL-1 β on cartilage (38). However, the biologic functions of endogenous IL-1 antagonists in OA tissues and their capacity to neutralize the increased level of active IL-1 β in situ remain unknown.

In OA, TNF α also appears to be an important mediator of matrix degradation and a pivotal cytokine in inducing synovial membrane inflammation. The proteolytic cleavage of the proform of this cytokine takes place at the cellular surface via a TNF α -converting enzyme named TACE, which belongs to a subfamily of proteases named adamalysin (39). An up-regulation of TACE gene expression in human OA cartilage has recently been reported (40). Once secreted, the cytokine protein oligomerizes to form trimers, which bind to 2 specific receptors (TNFR) on the cell membrane. These 2 TNFR (41,42) are named according to their molecular weight, TNFR55 and TNFR75. TNFR55 seems to be the dominant receptor responsible for mediating TNF α activity in OA chondrocytes and synovial cells. An enhanced expression of TNFR55 has been reported in these cells (43, 44).

Adding to the complexity of this cytokine is the recent finding that proteolytic cleavage of the extracellular domain of each TNFR produces sTNFR. The 2 soluble receptors sTNFR55 and sTNFR75 are produced spontaneously by OA synovial fibroblasts and chondrocytes (43,45). These cells have been found to release an increased amount of sTNFR75 (43,45), and an increased level of sTNFR has been found in the synovial fluid of patients with different forms of arthritis (46,47). It is believed that the biologic role of sTNFR varies depending on its concentration in the joint tissues. At low concentrations, sTNFR could stabilize the trimeric structure of TNF α , thereby increasing the half-life of bioactive TNF α . At high concentrations, sTNFR reduces the bioactivity of $TNF\alpha$ by competing for TNF binding with cell-associated receptors. Therefore, the low level of sTNFR found in OA tissues would be another factor favoring the catabolic effects of $TNF\alpha$.

Other proinflammatory cytokines, including IL-8, LIF, IL-6, IL-11, and IL-17, have been shown to be overexpressed in OA tissue, and should therefore be considered potential contributing factors in the pathogenesis of this disease. Two of them, IL-11 and IL-6, have also shown antiinflammatory properties.



Figure 2. Role of nitric oxide (NO) synthase in osteoarthritis. The up-regulation of the inducible form of nitric oxide synthase (iNOS) causes an excessive production of NO, which is responsible for inducing an inflammatory reaction, tissue destruction, as well as cell death.

Antiinflammatory cytokines and cytokine antagonists. A number of antiinflammatory cytokines, such as IL-4, IL-10, and IL-13, have been shown to be spontaneously elaborated by synovial membrane and cartilage, and are found in increased levels in the synovial fluid of OA patients (48). These cytokines exert their antiinflammatory properties through a number of mechanisms, resulting in a decrease in the production of IL-1 β , TNF α , and MMPs, up-regulation of IL-1 receptor antagonist (IL-1Ra) and tissue inhibitor of matrix metalloproteases 1 (TIMP-1), as well as inhibition of PGE₂ release (49–54).

IL-1Ra is a competitive inhibitor of IL-1R and can block a number of catabolic pathways related to OA, including PGE₂ synthesis, collagenase and NO production by chondrocytes, and cartilage matrix degradation. Even though a higher level of IL-1Ra is found in OA articular tissues, the ratio of IL-1Ra to IL-1 β is insufficient to deal with the increased level of IL-1 β found in those tissues (29,48).

Nitric oxide: a true catabolic factor. NO is a factor that is very likely involved in the promotion of cartilage catabolism in OA through a number of mechanisms (55) (Figure 2). OA cartilage produces a large amount of NO (56,57), and a high level of nitrites/ nitrates have been found in the synovial fluid and serum of arthritis patients (58), which is caused by an increased level of the inducible form of NO synthase (iNOS) (59,60). NO can inhibit the synthesis of cartilage matrix macromolecules such as aggrecans, can enhance MMP activity (61,62), and can reduce the synthesis of IL-1Ra by chondrocytes (57). The selective inhibition of iNOS has proven to exert positive effects on the progression of lesions in an experimental canine OA model (63).

Eicosanoids: prostaglandins and leukotrienes (Figure 3). The expression of the inducible cyclooxygenase, COX-2, is increased in OA chondrocytes that



Figure 3. Possible role of eicosanoid in osteoarthritis symptoms and pathophysiology. 5-LO = 5-lipoxygenase; COX-2 = cyclooxygenase 2; LTB₄ = leukotriene B₄; PGI₂ = prostaglandin I₂; PGE₂ = prostaglandin E₂; TXA₂ = thromboxane A₂.

spontaneously produce PGE_2 ex vivo (64). Findings in the literature on the effects of eicosanoid overproduction reveal a variety of both catabolic and anabolic activities. In part, this is due to the fact that different eicosanoid end-products (e.g., PGE_1 , PGE_2), acting via different PGE receptors and signaling pathways, have been shown to exert divergent effects on chondrocyte metabolism. Thus, the in vivo consequence of COX-2 overexpression in OA may lead to the production of a variety of prostanoids, of which the net effect on the disease process may be difficult to assess in vitro. However, because of the widespread, prolonged use of COX inhibitors in clinical practice, this is an area that merits further investigation, including the assessment of structural outcomes in the clinic (65).

Moreover, the role of products of the lipoxygenase pathway in OA is unclear at present. Leukotriene B_4 (LTB₄) activity was found to be elevated in the synovial fluid from patients with OA, and both LTB₄ and leukotriene C₄ production have been reported in OA synovial tissue, but not in chondrocytes (66–68). Although the leukotriene mechanism of action is not fully established, LTB₄ was reported to induce IL-1 β production in synovial cells (69,70). Since many of these products are produced in only minute amounts and are detected with difficulty by radioimmunoassay or enzyme-linked immunosorbent assay, this is a field that merits further investigation.

Chondrocyte apoptosis: an integral part of the disease process

Morphologic alterations in cartilage involve both extracellular matrix components and chondrocytes. Among the chondrocyte changes is cell cloning. Moreover, there is often an increased number of intracytoplasmic organelles reflecting the hypersynthetic state of these cells (71). There is also an increase in the number of cells exhibiting signs of degeneration or death, a phenomenon that has been shown to be related to both cell necrosis and apoptosis (programmed cell death). The latter involves a complex process related to the activation of several intracellular signaling pathways (72,73). Excess production of NO in OA tissues has been linked with cartilage chondrocyte apoptosis both in vitro and in vivo (74,75). The exact mechanism by which NO mediates apoptosis in OA chondrocytes is not yet completely understood. However, the activation of the caspase cascade seems to play an essential role.

Another possible mechanism that could also contribute to OA chondrocyte apoptosis has recently been identified. A subpopulation of OA chondrocytes (in superficial zones of the cartilage) expresses the Fas antigen, which upon ligand binding, could induce cell apoptosis (76). Interestingly, it is in that zone that most of the apoptotic cells are located. It is presently not known under which condition chondrocytes express Fas ligand, since its only possible source in the OA joint is inflammatory cells in the synovial tissue and fluid.

Inflammation: a therapeutic target

The main objectives in the management of OA are to reduce symptoms, minimize functional disability, and limit progression of the structural changes (77). Our

understanding of the role of catabolic factors in cartilage degradation and the implication of synovial inflammation and cytokines in disease progression has improved substantially in the last 2 decades (3,29). These findings have made possible more precise identification of pathways that have the potential to become therapeutic targets. These pathways can be modified to effectively retard the progression of the disease.

A number of such new agents, referred to as disease-modifying osteoarthritic drugs (DMOADs) (discussed below), are now the subject of preclinical and clinical trials. However, given these new insights, what are the implications for treatment with respect to currently available agents such as nonsteroidal antiinflammatory drugs (NSAIDs), acetaminophen, or intraarticular corticosteroids, or even the use of drugs such as methotrexate? Clinical studies to date have focused on the alleviation of signs and symptoms, for which comparable efficacy has generally been demonstrated for NSAIDs and acetaminophen in stable cohorts of patients with mild-to-moderate OA (78,79). Published data on intraarticular corticosteroids in OA have demonstrated short-term (up to 4 weeks) improvement of signs and symptoms compared with placebo (78,80). The comparative efficacy of these agents in the treatment of episodic crystal-induced inflammatory exacerbations superimposed on chronic OA in selected patients (81), in which one might predict superior efficacy for NSAIDs or corticosteroids over acetaminophen, has not been studied.

However, the current paradigm of inflammatory mediator production in OA focuses less on the induction of signs and symptoms (which has been studied extensively as noted above) than on the potential autocrine/ paracrine catabolic actions of these products on cartilage metabolism. Since there are in vitro data to indicate that corticosteroids inhibit synoviocyte and chondrocyte production of IL-1, COX-2, and TNF α (80), there has been speculation that intraarticular corticosteroid administration could exert a disease-modifying role in OA. However, a recent structural outcome study in humans has not been able to validate such a hypothesis (82). Similarly, despite speculation that NSAIDs exert beneficial or harmful effects on the integrity of articular cartilage, there are no validated imaging studies that shed light on this controversy. Predicting the net effects of COX inhibitors on cartilage structure is particularly difficult given the observed pleiotropic effects of individual eicosanoids in vitro on chondrocyte functions, as discussed above. Finally, studies of methotrexate on chondrocyte function, in vitro and in vivo, have failed to show significant effects on cartilage metabolism at clinically relevant concentrations (83).

Thus, although the long-term effects of available antiinflammatory agents on cartilage merit further investigation, there is also significant interest in new agents that have the potential to reduce or stop the progression of the structural changes observed in OA. Such agents offer great promise and are likely to lead to very significant changes in therapeutic approaches in the near future.

The different DMOAD agents presently in development or those targeting pathophysiologic processes and having therapeutic potential can be briefly summarized as follows.

Inhibitors of MMPs (Figure 1). As previously mentioned, some members of the MMP family are intimately involved in articular joint matrix degradation. These enzymes are synthesized as proenzymes and must be activated by proteolytic cleavage. A number of agents that bind the active site of the enzyme can inhibit its catalytic activity. Among these agents are natural MMP inhibitors such as TIMP. Increasing the local synthesis of TIMP would be an effective way to prevent connective tissue turnover and OA progression. However, this natural protein showed limitations with regard to its administration. Nonetheless, therapy with TIMP using recombinant protein and gene therapy has been shown to be effective in antimetastatic treatment (84). These findings, in turn, have generated a regain of interest in the use of TIMP as a therapeutic strategy for OA.

Strategies for the control of MMP synthesis/ activity, particularly using synthetic compounds, have been the focus of intensive research over the last decade (27). Although prospects for the prevention of cartilage macromolecule breakdown using synthetic MMP inhibitors look promising, opinions differ as to the best MMPs to target. Stopping the degradation of the collagen network is certainly logical, since it has been shown that its loss leads to irreversible damage. Therefore, collagenases are among the main candidates for inhibition. Collagenase 3 (MMP-13) seems to be a very attractive candidate, because it is the most potent proteolytic enzyme of the 3 collagenases for type II collagen and it is selectively expressed in pathologic conditions such as arthritis (85).

Antibiotics such as tetracycline and its semisynthetic forms (doxycycline and minocycline) have very significant inhibitory properties that impact MMP activity, even in vivo (86). Their main action is mediated by chelating the zinc present in the active site of MMPs. A potential additional therapeutic effect of the tetracyclines may be gained as a result of their capacity to inhibit the expression of iNOS and thereby block NO production (87). A clinical trial is presently underway to evaluate the therapeutic efficacy of doxycycline in patients with knee OA.

Inhibition of cytokine activity. As mentioned earlier, proinflammatory cytokines are predominant factors involved in the progression of OA. Among these, IL-1 β and TNF α play a pivotal role (30,31). Control of cytokine action can be modulated at different levels; therapeutic intervention could target the synthesis, the maturation, or the activity of those cytokines.

Antiinflammatory cytokines. Cytokines such as IL-4, IL-10, and IL-13 were demonstrated to effectively reduce the production of IL-1 β and TNF α in vitro while increasing the IL-1Ra production in OA synovium explants. These data suggest that these antiinflammatory cytokines could potentially be useful for the treatment of OA. So far, clinical trials have only evaluated the effects of IL-10 in RA patients and, as yet, no study is underway in OA patients.

IL-1 β and TNF α activity inhibition. As mentioned, both IL-1 β and TNF α are synthesized as inactive precursors and must be activated by an enzyme before being released extracellularly in their active forms, ICE and TACE, respectively. Therefore, the inhibition of IL-1 β and TNF α maturation by specific convertase inhibitors appears to be an attractive therapeutic target. In fact, it was recently shown that an ICE inhibitor can completely abrogate ex vivo the formation of active IL-1 β in OA tissue (34). In vivo, in an animal model, data showed that an ICE inhibitor effectively reduced the progression of murine type II collagen–induced arthritis (CIA) (88). The evaluation of the potential of ICE inhibition for the treatment of RA is presently underway.

Receptor blockade or molecular quenching. The IL-1 system is regulated by a natural antagonist of the receptor, namely IL-1Ra. In vivo experiments using intraarticular injections of IL-1Ra or IL-1Ra gene transfection were found to retard the progression of experimental OA (30,89,90). Based on these findings and results from clinical trials in RA patients, it is believed that the use of IL-1Ra for the treatment of OA holds promise. However, to our knowledge, no clinical trial is yet underway for the latter disease.

Another mechanism available to inhibit proinflammatory cytokines is the use of soluble receptors to neutralize the cytokine. Types I or II sIL-1R are potential therapeutic candidates. In human RA, the administration of sTNFR has been shown to be a very effective treatment (91). The role for TNF α in OA cartilage degradation is less clear than that of IL-1, although the production of both TNF α and its converting enzyme, TACE, are increased in OA (40). However, it is possible

Table 2. Gene therapy for osteoarthritis*

Potential targets	
Cartilage	
Catabolic factors (for example, MMPs, NO)	
Anabolic factors (growth factors)	
Apoptotic factors (for example, caspases, ceramides)	
Synovium	
Cytokines (for example, IL-1 β , TNF α)	
Antiinflammatory cytokines (IL-4, IL-10, IL-13)	
Cytokine receptor antagonist (IL-1Ra)	
Soluble receptors (sIL-1RII, sTNFR)	
Strategies	
Gene replacement	
Gene addition	
Gene control	

* In both cartilage and synovium, catabolic factors (for example, metalloproteases [MMPs], nitric oxide [NO]) and cytokines (for example, interleukin-1 β [IL-1 β], tumor necrosis factor α [TNF α]) should be either reduced or eliminated. In contrast, some growth factors and the cytokine receptor antagonist (interleukin-1Ra [IL-1Ra]) or soluble receptors (soluble IL-1 receptor type II [sIL-1RI], soluble tumor necrosis factor receptor [sTNFR]) should be stimulated.

that both IL-1 and TNF α contribute independently to articular degeneration. Depending on the availability of therapeutic agents with an acceptable risk:benefit ratio, trials that examine TNF α antagonism in OA could be considered.

Specific neutralizing antibodies against IL-1 or TNF α have been tested in different systems. The IL-1 antibody has been successfully tested in a CIA murine model of inflammatory arthritis (92). Treatment with an anti-TNF α antibody has also been shown to improve arthritis in an experimental model as well as RA in humans (93). No such treatment has yet been tested in OA.

Inhibition of intracellular signaling pathways. Several postreceptor signaling pathways are involved in the synthesis of cytokines. Two of these pathways, p38 mitogen-activated protein (MAP) kinase and nuclear factor κ B (NF- κ B), appear to be the major ones involved in mediating the synthesis of several inflammatory cytokines and MMPs and are likely to play a role in these pathways that are activated during the OA process (29).

Pyridinyl imidazole compounds that have the ability to inhibit p38 MAP kinase and block proinflammatory cytokine production have been named cytokinesuppressive antiinflammatory drugs, or CSAIDs. These compounds inhibit synthesis of proinflammatory cytokines such as IL-1 and TNF α at the translational level (94). They have proven therapeutic effectiveness in animal models of inflammatory arthritis (95). In addition, some CSAIDs were also shown to inhibit the production of NO by chondrocytes or by human OA cartilage (96,97).

Drugs that will target NF-kB activity/activation

could have definite potential for the treatment of arthritis. COX-2 and IL-1 β are but 2 of several genes modulated by NF- κ B activation. A recent report showed that specifically blocking the activation of this factor in vivo in the CIA model induced a marked reduction in the expression of IL-1 β and TNF α in synovium, as well as suppressing the degradation of bone and cartilage of the arthritic joint (98).

Inhibition of NO production. The discovery and characterization of the functions of the iNOS isoenzyme have provided the impetus for novel therapeutic approaches toward developing a potential new class of drugs. The challenge lies in making selective inhibitors that target only the inducible form of NOS in order not to down-regulate the constitutive OA physiologic isoform. The use of a selective iNOS inhibitor in a model of experimentally induced OA in dogs (99,100) was demonstrated to reduce in vivo the progression of early lesions, which was then associated with a reduction in cartilage MMP activity and IL-1 β in synovium. Moreover, it was shown that the selective inhibition of iNOS decreased in situ the level of chondrocyte apoptosis. These data bring forward the potential of selective iNOS inhibitors, not only as effective agents for the treatment of the signs and symptoms of OA, but also for diseasemodifying activity.

Antiapoptotic therapy. Chondrocyte apoptosis is a complex process mediated by the activation of several intercellular signaling pathways (74,75), including the caspase cascade which induces DNA damage (101). Current and future knowledge about its regulatory mechanisms will make it possible to develop a strategy for therapeutic approaches that could be targeted for future OA treatment. Targeting the caspase cascade or the mechanisms involved in caspase activation with the use of specific inhibitors is very appealing, although their potential side effects will require careful evaluation.

Gene therapy: an attractive concept and maybe more. Gene therapy (Table 2) in articular joint tissues can be used as a drug delivery system to modify or reestablish the balance between catabolic/anabolic factors or to modulate proinflammatory cytokines. Ideally, this must be done to the cell or must be tissue specific. The potential for the use of biologic molecules as therapeutic agents is limited. Lately, much attention has been focused on the use of gene transfer techniques. Their potential for the treatment of OA is of very significant interest, since a consistently high local concentration of the therapeutic protein in the joint can be provided and sustained delivery maintained over time. Several strategies to replace defective or deficient protein products are now under study (Table 2). Treatment approaches consist of various ex vivo or in vivo techniques using viral or nonviral vectors (102). One strategy consists of insertion into the cells of a gene enabling the production of a protein not normally expressed or expressed in low and insufficient amounts by the OA cells. The viral system is favored because it generally allows for a very effective transfer to a large percentage of cells while maintaining a sustained high level of protein expression that can be extended over significant periods of time. Ex vivo transfer of marker genes to OA cells has been demonstrated in experimental models with the use of a retroviral vector (89).

The selection or combination of the gene(s) that would offer the best protection against OA remains to be determined. The transfer of genes such as IL-1Ra, IL-10, and IL-13 has been studied using OA or inflammatory animal models (103). However, more specifically with regard to OA, the use of IL-1Ra gene therapy has elicited much attention. The rationale is based mainly on the fact that this antagonist has the ability in vitro to arrest cartilage degradation and in vivo to reduce the progression of experimental OA (30,89,90).

Conclusion

The current understanding of the pathophysiologic pathways involved in OA has evolved greatly in recent years. Specifically, the role of inflammation has been explored and new findings have allowed for a much better understanding of the disease process, the modulating factors, as well as the major regulators, which may have potential therapeutic value by specifically and effectively retarding the progression of this disease. A large amount of new information about OA pathophysiology and new targets for the development of therapeutic strategies has been generated from in vitro and experimental studies. Caution should obviously be exerted in extrapolating this to the clinical situation. Nevertheless, the future holds great promise for the development of new and successful approaches to the treatment of this disease.

REFERENCES

- Felson DT, Zhang Y, Hannan MT, Naimark A, Weissman B, Aliabadi P, et al. Risk factors for incident radiographic knee osteoarthritis in the elderly: the Framingham Study. Arthritis Rheum 1997;40:728–33.
- Williams CJ, Jimenez SA. Genetic and metabolic aspects. In: Reginster JY, Pelletier JP, Martel-Pelletier J, Henrotin Y, editors. Osteoarthritis: clinical and experimental aspects. Berlin: Springer-Verlag; 1999. p. 134–55.

- Pelletier JP, Martel-Pelletier J, Howell DS. Etiopathogenesis of osteoarthritis. In: Koopman WJ, editor. Arthritis & allied conditions: a textbook of rheumatology. 14th ed. Baltimore: Lippincott Williams & Wilkins; 2000. p. 2195–245.
- Nuki G. Role of mechanical factors in the aetiology, pathogenesis and progression of osteoarthritis. In: Reginster JY, Pelletier JP, Martel-Pelletier J, Henrotin Y, editors. Osteoarthritis: clinical and experimental aspects. Berlin: Springer-Verlag; 1999. p. 101–14.
- Haraoui B, Pelletier J-P, Cloutier J-M, Faure M-P, Martel-Pelletier J. Synovial membrane histology and immunopathology in rheumatoid arthritis and osteoarthritis: in vivo effects of antirheumatic drugs. Arthritis Rheum 1991;34:153–63.
- Farahat MN, Yanni G, Poston R, Panayi GS. Cytokine expression in synovial membranes of patients with rheumatoid arthritis and osteoarthritis. Ann Rheum Dis 1993;52:870–5.
- Smith MD, Triantafillou S, Parker A, Youssef PP, Coleman M. Synovial membrane inflammation and cytokine production in patients with early osteoarthritis. J Rheumatol 1997;24:365–71.
- Jiang Y, Peterfy CG, Zhao JJ, White DL, Lynch JA, Genant HK. Magnetic resonance imaging in osteoarthritis. In: Reginster J-Y, Pelletier J-P, Martel-Pelletier J, Henrotin Y, editors. Osteoarthritis: clinical and experimental aspects. Berlin: Springer-Verlag; 1999. p. 268–95.
- Sharif M, Saxne T, Shepstone L, Kirwan JR, Elson CJ, Heinegard D, et al. Relationship between serum cartilage oligomeric matrix protein levels and disease progression in osteoarthritis of the knee joint. Br J Rheumatol 1995;34:306–10.
- Clark AG, Jordan JM, Vilim V, Renner JB, Dragomir AD, Luta G, et al. Serum cartilage oligomeric matrix protein reflects osteoarthritis presence and severity: the Johnston County Osteoarthritis Project. Arthritis Rheum 1999;42:2356–64.
- Petersson IF, Boegard T, Svensson B, Heinegard D, Saxne T. Changes in cartilage and bone metabolism identified by serum markers in early osteoarthritis of the knee joint. Br J Rheumatol 1998;37:46–50.
- 12. Sharif M, Shepstone L, Elson CJ, Dieppe PA, Kirwan JR. Increased serum C reactive protein may reflect events that precede radiographic progression in osteoarthritis of the knee. Ann Rheum Dis 2000;59:71–4.
- Spector TD, Hart DJ, Nandra D, Doyle DV, Mackillop N, Gallimore JR, et al. Low-level increases in serum C-reactive protein are present in early osteoarthritis of the knee and predict progressive disease. Arthritis Rheum 1997;40:723–7.
- Goldberg RL, Huff JP, Lenz ME, Glickman P, Katz R, Thonar EJ-MA. Elevated plasma levels of hyaluronate in patients with osteoarthritis and rheumatoid arthritis. Arthritis Rheum 1991;34: 799–807.
- Otterness IG, Saltarelli MJ. Using molecular markers to monitor osteoarthritis. In: Tsokos GC, Moreland LW, Kammer GM, Pelletier J-P, Martel-Pelletier J, Gay S, editors. Modern therapeutics in rheumatic diseases. Totowa, NJ: Humana Press; in press.
- Kumon Y, Suehiro T, Nishiya K, Hashimoto K, Nakatani K, Sipe JD. Ferritin correlates with C-reactive protein and acute phase serum amyloid A in synovial fluid, but not in serum. Amyloid 1999;6:130–5.
- Belcher C, Yaqub R, Fawthrop F, Bayliss M, Doherty M. Synovial fluid chondroitin and keratan sulphate epitopes, glycosaminoglycans, and hyaluronan in arthritic and normal knees. Ann Rheum Dis 1997;56:299–307.
- Praest BM, Greiling H, Kock R. Assay of synovial fluid parameters: hyaluronan concentration as a potential marker for joint diseases. Clin Chim Acta 1997;266:117–28.
- Thonar EJ, Masuda K, Lenz ME, Hauselmann HJ, Kuettner KE, Manicourt DH. Serum markers of systemic disease processes in osteoarthritis. J Rheumatol Suppl 1995;43:68–70.

- Lohmander LS, Saxne T, Heinegard DK. Release of cartilage oligomeric matrix protein (COMP) into joint fluid after knee injury and in osteoarthritis. Ann Rheum Dis 1994;53:8–13.
- Verbruggen G, Veys EM. Numerical scoring systems for the anatomic evolution of osteoarthritis of the finger joints. Arthritis Rheum 1996;39:308–20.
- Cooke TD, Bennett EL, Ohno O. The deposition of immunoglobulins and complement in osteoarthritic cartilage. Int Orthop 1980;4:211–7.
- Myers SL, Brandt KD, Ehlich JW, Braunstein EM, Shelbourne KD, Heck DA, et al. Synovial inflammation in patients with early osteoarthritis of the knee. J Rheumatol 1990;17:1662–9.
- Lindblad S, Hedfors E. Arthroscopic and immunohistologic characterization of knee joint synovitis in osteoarthritis. Arthritis Rheum 1987;30:1081–8.
- 25. Krenn V, Hensel F, Kim HJ, Souto Carneiro MM, Starostik P, Ristow G, et al. Molecular IgV(H) analysis demonstrates highly somatic mutated B cells in synovialitis of osteoarthritis: a degenerative disease is associated with a specific, not locally generated immune response. Lab Invest 1999;79:1377–84.
- Nakamura H, Yoshino S, Kato T, Tsuruha J, Nishioka K. T-cell mediated inflammatory pathway in osteoarthritis. Osteoarthritis Cartilage 1999;7:401–2.
- Martel-Pelletier J, Tardif G, Fernandes JC, Pelletier J-P. Metalloproteases and their modulation as treatment in osteoarthritis. In: Tsokos GC, editor. Principles of molecular rheumatology. Totowa, NJ: Humana Press; 2000. p. 499–514.
- Attur MG, Patel IR, Patel RN, Abramson SB, Amin AR. Autocrine production of IL-1 beta by human osteoarthritisaffected cartilage and differential regulation of endogenous nitric oxide, IL-6, prostaglandin E2, and IL-8. Proc Assoc Am Physicians 1998;110:65–72.
- Martel-Pelletier J, di Battista JA, Lajeunesse D. Biochemical factors in joint articular tissue degradation in osteoarthritis. In: Reginster JY, Pelletier JP, Martel-Pelletier J, Henrotin Y, editors. Osteoarthritis: clinical and experimental aspects. Berlin: Springer-Verlag; 1999. p. 156–87.
- Caron JP, Fernandes JC, Martel-Pelletier J, Tardif G, Mineau F, Geng C, et al. Chondroprotective effect of intraarticular injections of interleukin-1 receptor antagonist in experimental osteoarthritis: suppression of collagenase-1 expression. Arthritis Rheum 1996;39:1535–44.
- 31. Van de Loo FAJ, Joosten LAB, van Lent PLEM, Arntz OJ, van den Berg WB. Role of interleukin-1, tumor necrosis factor α, and interleukin-6 in cartilage proteoglycan metabolism and destruction: effect of in situ blocking in murine antigen- and zymosan-induced arthritis. Arthritis Rheum 1995;38:164–72.
- Bertolini DR, Nedwin GE, Bringman TS, Smith DD, Mundy GR. Stimulation of bone resorption and inhibition of bone formation in vitro by human tumour necrosis factors. Nature 1986;319: 516–8.
- 33. Kronheim SR, Mumma A, Greenstreet T, Glackin PJ, van Ness K, March CJ, et al. Purification of interleukin-1 beta converting enzyme, the protease that cleaves the interleukin-1 beta precursor. Arch Biochem Biophys 1992;296:698–703.
- 34. Saha N, Moldovan F, Tardif G, Pelletier J-P, Cloutier J-M, Martel-Pelletier J. Interleukin-1β-converting enzyme/caspase-1 in human osteoarthritic tissues: localization and role in the maturation of interleukin-1β and interleukin-18. Arthritis Rheum 1999;42:1577–87.
- 35. Slack J, McMahan CJ, Waugh S, Schooley K, Spriggs MK, Sims JE, et al. Independent binding of interleukin-1 alpha and interleukin-1 beta to type I and type II interleukin-1 receptors. J Biol Chem 1993;268:2513–24.
- 36. Martel-Pelletier J, McCollum R, Di Battista J, Faure M-P, Chin JA, Fournier S, et al. The interleukin-1 receptor in normal and osteoarthritic human articular chondrocytes: identification as the

type I receptor and analysis of binding kinetics and biologic function. Arthritis Rheum 1992;35:530-40.

- 37. Sadouk M, Pelletier JP, Tardif G, Kiansa K, Cloutier JM, Martel-Pelletier J. Human synovial fibroblasts coexpress interleukin-1 receptor type I and type II mRNA: the increased level of the interleukin-1 receptor in osteoarthritic cells is related to an increased level of the type I receptor. Lab Invest 1995;73:347–55.
- Attur MG, Mandar D, Cipolletta C, Kang P, Goldring MB, Patel IR, et al. Reversal of autocrine and paracrine effects of interleukin-1 (IL-1) in human arthritis by type II IL-1 decoy receptor: potential for pharmacological intervention. J Biol Chem 2000; 275:40307–15.
- Black RA, Rauch CT, Kozlosky CJ, Peschon JJ, Slack JL, Wolfson MF, et al. A metalloproteinase disintegrin that releases tumour-necrosis factor-alpha from cells. Nature 1997;385: 729–33.
- Amin AR. Regulation of tumor necrosis factor-alpha and tumor necrosis factor converting enzyme in human osteoarthritis. Osteoarthritis Cartilage 1999;7:392–4.
- Loetscher H, Pan YCE, Lahm HW, Gentz R, Brockhaus M, Tabuchi H, et al. Molecular cloning and expression of the human 55 kd tumor necrosis factor receptor. Cell 1990;61:351–9.
- Schall TJ, Lewis M, Koller KJ, Lee A, Rice GC, Wong GH, et al. Molecular cloning and expression of a receptor for human tumor necrosis factor. Cell 1990;61:361–70.
- 43. Alaaeddine N, di Battista JA, Pelletier JP, Cloutier JM, Kiansa K, Dupuis M, et al. Osteoarthritic synovial fibroblasts possess an increased level of tumor necrosis factor-receptor 55 (TNF-R55) that mediates biological activation by TNF-alpha. J Rheumatol 1997;24:1985–94.
- Westacott CI, Atkins RM, Dieppe PA, Elson CJ. Tumour necrosis factor-alpha receptor expression on chondrocytes isolated from human articular cartilage. J Rheumatol 1994;21: 1710–5.
- 45. Brennan FM, Gibbons DL, Cope AP, Katsikis P, Maini RN, Feldmann M. TNF inhibitors are produced spontaneously by rheumatoid and osteoarthritic synovial joint cell cultures: evidence of feedback control of TNF action. Scand J Immunol 1995;42:158–65.
- Chikanza IC, Roux-Lombard P, Dayer JM, Panayi GS. Tumour necrosis factor soluble receptors behave as acute phase reactants following surgery in patients with rheumatoid arthritis, chronic osteomyelitis and osteoarthritis. Clin Exp Immunol 1993;92: 19–22.
- 47. Cope AP, Aderka D, Doherty M, Engelmann H, Gibbons D, Jones AC, et al. Increased levels of soluble tumor necrosis factor receptors in the sera and synovial fluid of patients with rheumatic diseases. Arthritis Rheum 1992;35:1160–9.
- Martel-Pelletier J, Alaaeddine N, Pelletier JP. Cytokines and their role in the pathophysiology of osteoarthritis. Front Biosci 1999;4:D694–703.
- Hart PH, Vitti GF, Burgess DR, Whitty GA, Piccoli DS, Hamilton JA. Potential antiinflammatory effects of interleukin 4: suppression of human monocyte tumor necrosis factor alpha, interleukin 1, and prostaglandin E₂. Proc Natl Acad Sci U S A 1989;86:3803–7.
- Essner R, Rhoades K, McBride WH, Morton DL, Economou JS. IL-4 down-regulates IL-1 and TNF gene expression in human monocytes. J Immunol 1989;142:3857–61.
- Vannier E, Miller LC, Dinarello CA. Coordinated antiinflammatory effects of interleukin 4: interleukin 4 suppresses interleukin 1 production but up-regulates gene expression and synthesis of interleukin 1 receptor antagonist. Proc Natl Acad Sci U S A 1992;89:4076–80.
- 52. Hart PH, Ahern MJ, Smith MD, Finlay-Jones JJ. Comparison of the suppressive effects of interleukin-10 and interleukin-4 on

synovial fluid macrophages and blood monocytes from patients with inflammatory arthritis. Immunology 1995;84:536-42.

- 53. Jovanovic D, Pelletier JP, Alaaeddine N, Mineau F, Geng C, Ranger P, et al. Effect of IL-13 on cytokines, cytokine receptors and inhibitors on human osteoarthritic synovium and synovial fibroblasts. Osteoarthritis Cartilage 1998;6:40–9.
- 54. Alaaeddine N, Di Battista JA, Pelletier J-P, Kiansa K, Cloutier J-M, Martel-Pelletier J. Inhibition of tumor necrosis factor α -induced prostaglandin E₂ production by the antiinflammatory cytokines interleukin-4, interleukin-10, and interleukin-13 in osteoarthritic synovial fibroblasts: distinct targeting in the signaling pathways. Arthritis Rheum 1999;42:710–8.
- 55. Amin AR, Abramson SB. The role of nitric oxide in articular cartilage breakdown in osteoarthritis. Curr Opin Rheumatol 1998;10:263–8.
- 56. Amin AR, di Cesare PE, Vyas P, Attur MG, Tzeng E, Billiar TR, et al. The expression and regulation of nitric oxide synthase in human osteoarthritis-affected chondrocytes: evidence for an inducible "neuronal-like" nitric oxide synthase. J Exp Med 1995; 182:2097–102.
- 57. Pelletier JP, Mineau F, Ranger P, Tardif G, Martel-Pelletier J. The increased synthesis of inducible nitric oxide inhibits IL-1Ra synthesis by human articular chondrocytes: possible role in osteoarthritic cartilage degradation. Osteoarthritis Cartilage 1996;4:77–84.
- 58. Farrell AJ, Blake DR, Palmer RM, Moncada S. Increased concentrations of nitrite in synovial fluid and serum samples suggest increased nitric oxide synthesis in rheumatic diseases. Ann Rheum Dis 1992;51:1219–22.
- McInnes IB, Leung BP, Field M, Wei XQ, Huang FP, Sturrock RD, et al. Production of nitric oxide in the synovial membrane of rheumatoid and osteoarthritis patients. J Exp Med 1996;184: 1519–24.
- 60. Grabowski PS, Wright PK, Van't Hof RJ, Helfrich MH, Ohshima H, Ralston SH. Immunolocalization of inducible nitric oxide synthase in synovium and cartilage in rheumatoid arthritis and osteoarthritis. Br J Rheumatol 1997;36:651–5.
- Taskiran D, Stefanovic-Racic M, Georgescu HI, Evans CH. Nitric oxide mediates suppression of cartilage proteoglycan synthesis by interleukin-1. Biochem Biophys Res Commun 1994;200: 142–8.
- 62. Murrell GAC, Jang D, Williams RJ. Nitric oxide activates metalloprotease enzymes in articular cartilage. Biochem Biophys Res Commun 1995;206:15–21.
- Pelletier J-P, Jovanovic D, Fernandes JC, Manning P, Connor JR, Currie MG, et al. Reduced progression of experimental osteoarthritis in vivo by selective inhibition of inducible nitric oxide synthase. Arthritis Rheum 1998;41:1275–86.
- 64. Amin AR, Attur MG, Patel RN, Thakker GD, Marshall PJ, Rediske J, et al. Superinduction of cyclooxygenase-2 activity in human osteoarthritis-affected cartilage: influence of nitric oxide. J Clin Invest 1997;99:1231–7.
- 65. Abramson SB. The role of COX-2 produced by cartilage in arthritis. Osteoarthritis Cartilage 1999;7:380-1.
- 66. Wittenberg RH, Willburger RE, Kleemeyer KS, Peskar BA. In vitro release of prostaglandins and leukotrienes from synovial tissue, cartilage, and bone in degenerative joint diseases. Arthritis Rheum 1993;36:1444–50.
- Amat M, Díaz C, Vila L. Leukotriene A₄ hydrolase and leukotriene C₄ synthase activities in human chondrocytes: transcellular biosynthesis of leukotrienes during granulocyte–chondrocyte interaction. Arthritis Rheum 1998;41:1645–51.
- 68. Atik OS. Leukotriene B_4 and prostaglandin E_2 -like activity in synovial fluid in osteoarthritis. Prostaglandins Leukot Essent Fatty Acids 1990;39:253–4.
- 69. Rainsford KD, Ying C, Smith F. Effects of 5-lipoxygenase inhibitors on interleukin production by human synovial tissues in

organ culture: comparison with interleukin-1-synthesis inhibitors. J Pharm Pharmacol 1996;48:46–52.

- 70. Kageyama Y, Koide Y, Miyamoto S, Yoshida TO, Inoue T. Leukotrien B_4 -induced interleukin-1 β in synovial cells from patients with rheumatoid arthritis. Scand J Rheumatol 1994;23: 148–50.
- Pelletier J-P, Martel-Pelletier J, Altman RD, Ghandur-Mnaymneh L, Howell DS, Woessner JF Jr. Collagenolytic activity and collagen matrix breakdown of the articular cartilage in the Pond-Nuki dog model of osteoarthritis. Arthritis Rheum 1983; 26:866–74.
- Blanco FJ, Guitian R, Vázquez-Martul E, de Toro FJ, Galdo F. Osteoarthritis chondrocytes die by apoptosis: a possible pathway for osteoarthritis pathology. Arthritis Rheum 1998;41:284–9.
- Vaishnaw AK, McNally JD, Elkon KB. Apoptosis in the rheumatic diseases. Arthritis Rheum 1997;40:1917–27.
- Hashimoto S, Takahashi K, Amiel D, Coutts RD, Lotz M. Chondrocyte apoptosis and nitric oxide production during experimentally induced osteoarthritis. Arthritis Rheum 1998;41: 1266–74.
- Blanco FJ, Ochs RL, Schwarz H, Lotz M. Chondrocyte apoptosis induced by nitric oxide. Am J Pathol 1995;146:75–85.
- Hashimoto S, Setareh M, Ochs RL, Lotz M. Fas/Fas ligand expression and induction of apoptosis in chondrocytes. Arthritis Rheum 1997;40:1749–55.
- Pelletier JP, Choquette D, Haraoui B, Raynauld JP, Rich E, Fernandes JC, et al. Pharmacologic therapy of osteoarthritis. Curr Rheumatol Rep 1999;1:54–8.
- Towheed TE, Hochberg MC. A systematic review of randomized controlled trials of pharmacological therapy in osteoarthritis of the knee, with an emphasis on trial methodology. Semin Arthritis Rheum 1997;26:755–70.
- American College of Rheumatology Subcommittee on Osteoarthritis Guidelines. Recommendations for the medical management of osteoarthritis of the hip and knee: 2000 update. Arthritis Rheum 2000;43:1905–15.
- Creamer P. Intra-articular corticosteroid treatment in osteoarthritis. Curr Opin Rheumatol 1999;11:417–21.
- McCarthy GM. Role of crystal deposition in the osteoarthritic joint. In: Reginster JY, Pelletier JP, Martel-Pelletier J, Henrotin Y, editors. Osteoarthritis: clinical and experimental aspects. Berlin: Springer-Verlag; 1999. p. 210–27.
- Raynauld JP, Buckland-Wright JC, Tremblay JL, Khy V, Bertrand C, Choquette D, et al. Clinical trials: impact of intraarticular steroid injections on the progression of knee osteoarthritis [abstract]. Osteoarthritis Cartilage 2000;8:S16.
- Neidel J, Sova L, Schroers B, Sintermann F, Manzke O, Bohlen H. Effects of methotrexate on normal articular cartilage in vitro and in vivo. Ann Rheum Dis 1998;57:414–21.
- Wojtowicz-Praga SM, Dickson RB, Hawkins MJ. Matrix metalloproteinase inhibitors. Invest New Drugs 1997;15:61–75.
- Martel-Pelletier J, Pelletier JP. Wanted—the collagenase responsible for the destruction of the collagen network in human cartilage! Br J Rheumatol 1996;35:818–20.
- Yu LP Jr, Smith GN Jr, Brandt KD, Myers SL, O'Connor BL, Brandt DA. Reduction of the severity of canine osteoarthritis by prophylactic treatment with oral doxycycline. Arthritis Rheum 1992;35:1150–9.
- Attur MG, Patel RN, Patel PD, Abramson SB, Amin AR. Tetracycline up-regulates COX-2 expression and prostaglandin E2 production independent of its effect on nitric oxide. J Immunol 1999;162:3160–7.
- 88. Ku G, Faust T, Lauffer LL, Livingston DJ, Harding MW. Interleukin-1 β converting enzyme inhibition blocks progression

of the type II collagen-induced arthritis in mice. Cytokine 1996; 8:377-86.

- 89. Pelletier J-P, Caron JP, Evans C, Robbins PD, Georgescu HI, Jovanovic D, et al. In vivo suppression of early experimental osteoarthritis by interleukin-1 receptor antagonist using gene therapy. Arthritis Rheum 1997;40:1012–9.
- Fernandes JC, Tardif G, Martel-Pelletier J, Lascau-Coman V, Dupuis M, Moldovan F, et al. In vivo transfer of interleukin-1 receptor antagonist gene in osteoarthritic rabbit knee joints: prevention of osteoarthritis progression. Am J Pathol 1999;154: 1159–69.
- Franklin CM. Clinical experience with soluble TNF p75 receptor in rheumatoid arthritis. Semin Arthritis Rheum 1999;29:172–81.
- 92. Joosten LAB, Helsen MMA, van de Loo FAJ, van den Berg WB. Anticytokine treatment of established type II collagen–induced arthritis in DBA/1 mice: a comparative study using anti-TNF α , anti–IL-1 α/β , and IL-1Ra. Arthritis Rheum 1996;39:797–809.
- 93. Maini R, St Clair EW, Breedveld F, Furst D, Kalden J, Weisman M, et al. Infliximab (chimeric anti-tumour necrosis factor alpha monoclonal antibody) versus placebo in rheumatoid arthritis patients receiving concomitant methotrexate: a randomised phase III trial. ATTRACT Study Group. Lancet 1999;354: 1932–9.
- 94. Young P, McDonnell P, Dunnington D, Hand A, Laydon J, Lee J. Pyridinyl imidazoles inhibit IL-1 and TNF production at the protein level. Agents Actions 1993;39 Spec No:C67–9.
- 95. Badger AM, Griswold DE, Kapadia R, Blake S, Swift BA, Hoffman SJ, et al. Disease-modifying activity of SB 242235, a selective inhibitor of p38 mitogen-activated protein kinase, in rat adjuvant-induced arthritis. Arthritis Rheum 2000;43:175–83.
- Badger AM, Cook MN, Lark MW, Newman-Tarr TM, Swift BA, Nelson AH, et al. SB 203580 inhibits p38 mitogen-activated protein kinase, nitric oxide production, and inducible nitric oxide synthase in bovine cartilage-derived chondrocytes. J Immunol 1998;161:467–73.
- 97. Martel-Pelletier J, Mineau F, Jovanovic D, Di Battista JA, Pelletier J-P. Mitogen-activated protein kinase and nuclear factor κB together regulate interleukin-17–induced nitric oxide production in human osteoarthritic chondrocytes: possible role of transactivating factor mitogen–activated protein kinase–activated protein kinase (MAPKAPK). Arthritis Rheum 1999;42:2399–409.
- 98. Tomita T, Takeuchi E, Tomita N, Morishita R, Kaneko M, Yamamoto K, et al. Suppressed severity of collagen-induced arthritis by in vivo transfection of nuclear factor κB decoy oligodeoxynucleotides as a gene therapy. Arthritis Rheum 1999; 42:2532–42.
- Pelletier JP, Lascau-Coman V, Jovanovic D, Fernandes JC, Manning P, Currie MG, et al. Selective inhibition of inducible nitric oxide synthase in experimental osteoarthritis is associated with reduction in tissue levels of catabolic factors. J Rheumatol 1999;26:2002–14.
- 100. Pelletier J-P, Jovanovic DV, Lascau-Coman V, Fernandes JC, Manning PT, Connor JR, et al. Selective inhibition of inducible nitric oxide synthase reduces progression of experimental osteoarthritis in vivo: possible link with the reduction in chondrocyte apoptosis and caspase 3 level. Arthritis Rheum 2000;43:1290–9.
- Thornberry N, Lazebnik YA. Caspases: enemies within. Science 1998;281:1312–6.
- Evans CH, Robbins PD. Gene therapy for arthritis. In: Wolff JA, editor. Gene therapeutics: methods and applications of direct gene transfer. Boston: Birkhauser; 1994. p. 320–43.
- Fernandes JC, Martel-Pelletier J, Pelletier JP. Gene therapy for osteoarthritis: new perspectives for the twenty-first century. Clin Orthop 2000;379 Suppl:S262–72.